Pathogenesis and treatment of non-alcoholic steatohepatitis and its fibrosis

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Running title
NAFLD mechanism and therapy

Abbreviations
NASH: Non-alcoholic steatohepatitis; RAS: Renin-angiotensin system; NAFLD: Non-alcoholic fatty liver disease; SFA: saturated fatty acid; DNL: De novo lipogenesis; TG: Triglyceride; FFA: Free fatty acid; ER: endoplasmic reticulum; HSC: Hepatic stellate cell; TGF-β: transforming growth factor β; SCFA: Short-chain fatty acid

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ABSTRACT
The initial presentation of non-alcoholic steatohepatitis (NASH) is hepatic steatosis. The dysfunction of lipid metabolism within hepatocytes caused by genetic factors, diet, and insulin resistance causes lipid accumulation. Lipotoxicity, oxidative stress, mitochondrial dysfunction, and endoplasmic reticulum stress would further contribute to hepatocyte injury and death, leading to inflammation and immune dysfunction in the liver. During the healing process, the accumulation of an excessive amount of fibrosis might occur while healing. During the development of NASH and liver fibrosis, the gut-liver axis, adipose-liver axis, and renin-angiotensin system (RAS) may be dysregulated and impaired. Translocation of bacteria or its end-products entering the liver could activate hepatocytes, Kupffer cells, and hepatic stellate cells, exacerbating hepatic steatosis, inflammation, and fibrosis. Bile acids regulate glucose and lipid metabolism through Farnesoid X receptors in the liver and intestine. Increased adipose tissue-derived non-esterified fatty acids would aggravate hepatic steatosis. Increased leptin also plays a role in hepatic fibrogenesis, and decreased adiponectin may contribute to hepatic insulin resistance. Moreover, dysregulation of peroxisome proliferator-activated receptors in the liver, adipose, and muscle tissues may impair lipid metabolism. Moreover, the RAS may contribute to hepatic fatty acid metabolism, inflammation, and fibrosis. The treatment includes lifestyle modification, pharmacological therapy, and non-pharmacological therapy. Currently, weight reduction by lifestyle modification or surgery is the most effective therapy. However, vitamin E, pioglitazone, and obeticholic acid have been suggested. In this review, we will introduce some new clinical trials and experimental therapies for the treatment of NASH and related fibrosis.

Keywords
Non-alcoholic fatty liver disease, lipotoxicity, chronic liver injury, mechanism, therapy
INTRODUCTION

Non-alcoholic fatty liver disease (NAFLD) is currently the most prevalent liver disease worldwide. NAFLD is a wide hepatic spectrum, ranging from simple steatosis to non-alcoholic steatohepatitis (NASH), which leads to progressive fibrosis, cirrhosis, and hepatocellular carcinoma [1]. Fat accumulation in hepatocytes sensitizes hepatocytes to injury, leading to cell death, inflammatory cells recruitment and activation of hepatic stellate cells (HSCs) [2]. The pathogenesis of NASH and its fibrosis has been broadly investigated for decades and the development and progression of NASH and liver fibrosis involves complex interplay of numerous determinants. Understanding of the pathogenesis of NASH and liver fibrosis is important for the diagnosis and development of treatment. Although new drugs have been developed to target liver inflammation and fibrosis in NASH, only a minority of patients achieve treatment response [3]. Thus, there is still an urgent need to develop new therapeutic agents for NASH.

PATHOGENESIS OF NASH

Development of hepatic steatosis

Diet

High-fat diet can result in hepatic steatosis in humans. Liver fat increased by 35% in overweight nondiabetic women after a 2-week isocaloric high-fat diet (56% total energy from fat) [4]. A 3-day high-fat, high-energy diet in healthy males resulted in major increases in plasma triglyceride (TG) and non-esterified fatty acid (NEFA) concentrations and hepatic TG [5]. A single energy-dense, high-fat meal induced net lipid accumulation in the liver of healthy subjects [6]. Moreover, palm oil administration in lean, healthy individuals decreased whole-body, hepatic, and adipose tissue insulin sensitivity by 25%, 15%, and 34%, respectively, increased hepatic triglyceride and ATP content by 35% and 16%, respectively, increased hepatic gluconeogenesis by 70%, and declined glycogenolysis by 20% [7]. In
young Finnish adults, serum fatty acid saturation independently predicted 10-year risk for fatty liver and omega-6 (ω6) fatty acids inversely associated with fatty liver [8]. A long-term hypercaloric diet, rich in SFA, showed a marked increased liver fat content by 50% and ω6 polyunsaturated fatty acids (PUFA) decreased fatty liver in overweight humans [9]. However, a lipidomic analysis showed that the n-6: n-3 free fatty acids (FFAs) ratio increased in NASH livers as compared to normal livers [10]. These studies suggest that hypercaloric diet, especially fat and sugar contribute to fatty liver; SFA and fructose are more detrimental but the role of the ω6/ω3 fat ratio is controversial.

**Physical inactivity**

NAFLD patients have low level of physical activity than normal controls. Gerber et al. showed that the average physical activity, counted by an accelerometer of NAFLD subjects, was about 28.7 counts/minute/day [11]. In an Asian group, prolonged sitting time and decreased physical activity level were found to positively associated with the prevalence of NAFLD and these associations were also observed in subjects with BMI <23 kg/m² [12]. However, the detail mechanism of sedentary behavior or low physical activity leading to fatty liver remains unclear. Lower expenditure of energy or lower skeletal muscle mass might explain a possible connection between sedentary behavior and NAFLD.

**Insulin resistance**

NAFLD is strongly associated with reduced whole body insulin sensitivity, increased hepatic and adipose tissue insulin resistance [13, 14]. Insulin resistance can lead to hepatic fat accumulation by increasing FFA delivery to the liver, increasing DNL and decreasing hepatic fatty acid oxidation. A landmark study performed by Donelly et al. demonstrated that in NAFLD patients, about 59% liver triacylglycerol arose from NEFAs, 26.1% from DNL and 14.9% from the diet and the liver demonstrated reciprocal use of adipose and dietary fatty
acids. DNL was elevated in the fasting state without diurnal variation [15]. Insulin resistance can impair the insulin suppression of lipolysis of peripheral adipose tissues, leading to increased delivery of FFAs to the liver [16]. Insulin can stimulate sterol receptor binding protein 1-c (SREBP1c), increasing DNL in the liver [17, 18]. Chronic hyperinsulinemia results in the cytoplasmic localization and inactivation of Foxa2 phosphorylation in hepatocytes, thereby promoting lipid accumulation and insulin resistance in the liver [19].

**Genetic factor**

There are several gene variants associated with NAFLD and NASH. The first fatty liver gene identified by Romeo et al. is patatin-like phospholipase domain-containing 3 (PNPLA3) [20]. The single nucleotide polymorphism (SNP) rs738409 causes the missense sequence variation I148M, impairing the phospholipase activity and increasing hepatic fat content [20].

Glucokinase regulatory protein (GCKR) can regulate hepatic glucose uptake and hepatic glucokinase activity, and the intronic SNP rs780094 is associated with hepatic lipid content [21, 22]. The SNP rs1260326 (C > T; P446L), GCKRP446L can decrease the inhibition of glucokinase, leading to increased glycolytic flux to hepatocytes, then hepatic steatosis [23]. The rs58542926 (G > A; E167K) variant, transmembrane 6 superfamily 2 (TM6SF2), was associated with increased hepatic triglyceride content [24]. Inhibition of TM6SF2 in hepatocytes reduced the secretion of VLDL, leading to the retention of triglycerides [25]. In a Taiwanese population, a variant in the immunity-related GTPase M (IRGM) gene (rs10065172 TT genotype) independently increased the odds ratio of NAFLD by 2.04 by altering hepatic lipid metabolism through the autophagy pathway [26]. Similarly, the IRGM rs10065172 variant increases the risk for hepatic steatosis but not for liver inflammation or fibrosis in obese Italian children [27]. Recently, mechanisms underlying metabolic and genetic components of NAFLD are found to be fundamentally different in patients. The metabolic component is characterized by hepatic oversupply of sugars and lipids, while the
genetic component is characterized by impaired hepatic mitochondrial function, reducing the liver’s ability to metabolize these substrates [28].

**Epigenetic factor**

Using epigenome-wide association study in peripheral blood cells, 22 CpGs were found to be associated with hepatic fat in European participants; 19 CpGs were annotated to 18 unique genes upregulated in the liver, including DHCR24, SLC43A1, CPT1A, SREBF1, SC4MOL, and SLC9A3R1 [29]. Some alternations of intrahepatic miRNA have been associated with hepatic steatosis. The serum levels of miR-122 and miR-192 were upregulated in patients with simple steatosis than in normal controls [30] Administration of exosomes transfected with obesity-associated miRNA induced hepatic steatosis in lean mice [31]. miR-122 inhibition in normal mice caused increased hepatic fatty acid oxidation [32]. Decreased miR-122-5p in the human liver was associated with impaired fatty acid usage [33]. However, deletion of mouse miR-122 resulted in hepatosteatosis, inflammation and the development of tumors [34]. The expression of miR-34 was elevated in NAFLD patients. miR-34a down-regulated autophagy in hepatocytes by targeting ATG4B and Rab-8B and suppressed mitochondrial biogenesis, leading to lipids accumulation in the liver [35].

**Lipotoxicity**

**ER stress**

The endoplasmic reticulum (ER) is responsible for protein folding, and the accumulation of misfolded or unfolded proteins leads to stress and the activation of the unfolded protein response (UPR) [36]. There are three sensor proteins that activate UPR, namely the inositol-requiring enzyme 1 (IRE1), the protein kinase R (double-stranded RNA-activated protein kinase)-like ER kinases (PERK), and the activating transcription factor 6 (ATF6). The UPR can cause inflammation, inflammasome activation, and death of hepatocytes [37]. Patients
with NASH were specifically associated with failure to generate X-box-binding protein 1 (XBP-1) protein and activation of JNK [38]. Palmitate can induce the ER stress response as demonstrated by increase in C/EBP homologous protein (CHOP) expression, eIF2-alpha phosphorylation, XBP-1 splicing, and JNK activation with increased expression of the BH3-only proteins PUMA and Bim [39]. Perturbation of membrane lipid composition could promote IRE1 and PERK activation, suggesting a lipid-sensing mechanism for ER sensors to activate the UPR [40]. NFATc1 drives hepatocyte damage and inflammation through activation of the PERK-CHOP [41].

**Mitochondrial dysfunction**

Increased hepatic fat would increase hepatic fat oxidation with increased mitochondrial respiration [42, 43]; however, decreased efficiency of respiratory chain complexes with greater mitochondrial uncoupling and leaking activity was found in patients with NAFLD [43, 44]. Chronic mitochondrial dysfunction in the state of lipid overload led to excessive leakage of electrons from mitochondrial respiratory complexes, leading to oxidative stress [45]. Voltage-dependent anion channel acted as an early sensor of lipid toxicity and its glycogen synthase kinase 3-mediated phosphorylation status controlled outer mitochondrial membrane permeabilization in hepatocytes with fat accumulation [46]. Exposure of hepatocytes to saturated FFAs caused mitochondrial depolarization, cytochrome c release, and increased ROS production [47]. Furthermore, intake of SFAs can affect the composition of mitochondrial membrane and decrease the efficiency of the respiratory transport chain, resulting in increased oxidative stress and chronic liver injury [48]. Peng *et al.* had found that hepatic cardiolipin and ubiquinone accumulated in NAFL, and levels of acylcarnitine increased with NASH and proposed that increased levels of cardiolipin and ubiquinone may help to preserve mitochondrial function in early NAFLD; however, mitochondrial function eventually fails with progression of NASH, leading to increased acylcarnitine [49]. Moreover,
SFAs increased ceramide synthesis in hepatocytes [50], which correlated with hepatocyte death via mitochondrial failure [51, 52].

**Lysosomal dysfunction**

It has been shown that hepatic activity of lysosomal acid lipase and lysosomal acidification, markers of lysosomal dysfunction, are decreased in patients with NAFLD [53, 54]. Both steatotic- and asparagine-treated hepatocytes showed reduced lysosomal acidity and retention of lysosomal calcium [55]. FFAs resulted in Bax translocation to lysosomes and lysosomal destabilization with release of cathepsin B into the cytosol, leading to nuclear factor kappa B-dependent tumor necrosis factor alpha expression and apoptosis [56, 57]. Lysosomal permeabilization and cathepsin B redistribution into the cytoplasm occurred several hours prior to mitochondrial dysfunction [47]. Furthermore, autophagy, which delivered intracellular proteins and organelles sequestered in double-membrane vesicles (autophagosomes) to lysosomes for degradation and use as an energy source. Autophagy in regulating intracellular lipid stores is called macrolipophagy [58]. Toxic fatty acids inhibited autophagic flux with reduction in lipophagy, which could lead to cell injury [59].

**Oxidative stress and apoptosis**

The main mechanisms of fatty acid-induced damage are oxidative stress and increased pro-inflammatory cytokines [2]. These insults from the ER stress, mitochondrial dysfunction, oxidative stress in the hepatocytes after lipid accumulation could cause lipotoxicity, leading to apoptosis, necroptosis, or pyroptosis [60, 61]. Saturated FFAs can induce apoptosis through intrinsic and extrinsic pathways. The oxidative stress and ER stress induced by accumulated fatty acids can activate CHOP and JNK, then upregulate Bim, Bax, and Bak, leading to release of cytochrome C and caspase 9-associated apoptosis. In addition, death receptor pathways including TRAIL/TRAILR, TNFa/TNFR1, and Fas ligand/Fas were noted to be
activated by FFAs on hepatocytes [62].

**DEVELOPMENT OF NASH FIBROSIS**

Liver fibrosis is the most important risk factor for liver cancer in patients with NAFLD and decompensated cirrhosis [63]. In patients with NAFLD, age and comorbidities including hypertension, overweighted, and diabetes mellitus are risk factors for progression of fibrosis [64-66].

Lipotoxic damage in hepatocytes would release cytokines and chemokines, then activate innate and adaptive immune cells including macrophages, dendritic cells, lymphocytes, and neutrophils, leading to an inflammation cascade [67]. Damaged hepatocytes also release extracellular vesicles containing exosomes, microparticles and apoptotic bodies. These vesicles, containing signaling proteins, sonic hedgehog, lipids, mRNAs, non-coding RNAs, and DNA, can induce inflammation, fibrosis by activating non-parenchymal cells, and recruitment of immune cells [68, 69]. Meanwhile, apoptotic bodies can also be engulfed by stellate cells and subsequently induce HSC activation which increases expression of α-smooth muscle actin, transforming growth factor β (TGF-β), and collagen type I [70]. Moreover, hedgehog (Hh) pathway was not only activated in hepatocytes, leading to macrophage recruitment and progression of inflammation [71], but it also induced epithelial-to-mesenchymal transitions (EMT) in ductular-type progenitors [72]. Cholangiocytes and natural killer T cells also activated hedgehog-osteopontin pathway and promoted fibrogenic responses of HSCs in NASH [73, 74].

Toxic fatty acids were able to directly affect Kupffer cells (KCs) and HSCs, which may contribute to the activation of inflammation and fibrosis. Palmitic acids activated toll-like receptor (TLR)2 and TLR4 in macrophages with induction of inflammatory signaling [75]. KCs exhibited a pro-inflammatory response with elevated levels of TNFα,
IL6, and IL1β after treatment by palmitic acids [75]. Palmitate induced ER stress and actin stress fiber formation in activated HSCs. Oleate induced the inflammatory signal and decreased cytoskeleton proteins in activated HSCs [76]. Free cholesterol was increased in patients with NAFLD and the accumulation of free cholesterol in HSCs sensitized these cells to TGFβ-induced activation, leading to exaggerated liver fibrosis in NASH [10, 77].

Insulin exerts profibrogenic activity. Insulin itself induces HSC mitogenesis and collagen synthesis [78, 79]. However, insulin enhances the expression of smooth muscle actin-α in quiescent but not in activated HSC through the PI3K/Akt-p70S6K pathway [80].

HSCs express PNPLA3 and membrane-bound O-acyltransferase domain-containing protein 7 (MBOAT7) [81, 82]. Increased PNPLA3 expression reduced lipid droplet content in HSCs [81]. Autophagy promotes loss of lipids in HSCs to provide energy for HSC activation [83]. PNPLA3^I148M can interfere with retinol production and release of HSCs by affecting retinyl-palmitate lipase activity, which may promote fibrosis progression [81]. The MBOAT7 rs641738 T allele was associated with lower protein expression in the liver and changes in plasma phosphatidylinositol species were consistent with decreased MBOAT7 function [82]. Hepatocyte-specific knockout of Mboat7 increased hepatic fibrosis with increased total lysophosphatidylinositol levels [84], which could promote the initiation of HSC activation by stimulating G-protein receptor 55 [85]. TM6SF2^{E167K} was associated with higher risk of advanced fibrosis in NAFLD patients [86]. Furthermore, the gene encoding for the hepatic hydroxysteroid 17-beta dehydrogenase 13 (HSD17B13) linked hepatic phospholipids and chronic liver injury in NAFLD patients [87-89]. The HSD17B13 rs72613567 variant led to loss of enzyme function, contributing to reduced inflammation and fibrosis in the liver [88]. In addition, the HSD17B13 rs72613567 variant affected retinol metabolism by reducing activity of retinyl-palmitate lipase, mediating antifibrotic and anti-inflammation effects [90].
Hypomethylation or hypermethylation of genes involved in the wound-healing process in NAFLD could be used to distinguish between patients with mild fibrosis from those with severe fibrosis in NAFLD. Hypermethylation at specific CpGs within TGFβ1 and PDGF, and hypomethylation at specific CpGs within peroxisome proliferator-activated receptor (PPAR) α and PPARδ in patients with mild fibrosis were found [91].

**ORGAN-ORGAN INTERACTION LEADING TO PROGRESSION OF NASH AND ITS FIBROSIS**

**Gut-liver axis**

Compared with healthy adults, patients with NAFLD had a higher proportion of Firmicutes in the intestine and the relative numbers of Bacteroidetes, Enterobacteriaceae and Ruminococcaceae families were reduced [92, 93]. Dysbiosis may disturb gut barriers, and bacteria and its products from the gut, such as endotoxin and cytokines, that promote inflammation, could enter the liver through blood, and activate the immune response in the liver and increase liver inflammation and fibrosis [94, 95]. Compared with healthy people, patients with NAFLD had dysbiosis and increased intestinal permeability, and patients with steatohepatitis were observed to have endotoxemia. [96, 97] Low-dose endotoxin stimulations were able to produce steatohepatitis in obese mice [98]. Conversely, blocking the signals caused by the immune system to recognize bacteria and its products effectively improved the severity of steatohepatitis [99, 100]. Bacterial products and translocated lipopolysaccharide stimulated the hepatic innate immune system through TLR 4 signaling, predominantly on HSCs and Kupffer cells (KCs) [101]. TLR4-mediated stimulation of HSCs led to HSC activation and KC activation [102]. In turn, KCs produced TGF-β, stimulating fibrogenesis, and the proinflammatory cytokines, propagating hepatic inflammation. KCs also produce reactive oxygen species, leading to generation of other reactive nitrogen species and local tissue damage [102, 103]. Mice fed with high fat diet for only 1 week undergo a diet-induced dysbiosis, driving
the damage on gut vascular barrier and causing bacterial translocation into the liver [104]. However, only 42.1% of patients with steatohepatitis had elevated endotoxin levels [105] and 39.1% of fatty liver patients had increased intestinal permeability [106]. Therefore, bacterial translocation due to gut barrier impairment may play a partial role in the development and progression of NAFLD and its fibrosis.

Some metabolites in blood and feces have been found to rely on bacterial synthesis, including choline and choline-related metabolites, bile acids, short-chain fatty acids (SCFAs), and ethanol, which may contribute to the pathogenesis of fatty liver. In animal experiments, the gut microbiota of mice fed with high-fat diet could convert choline into trimethylamine, reduce the bioavailability of choline, and produce a phenomenon similar to choline-deficient diet, leading to decreased excretion of VLDL from liver cells and increased liver fat accumulation [107]. Intestinal dysbiosis would increase the deoxycholic acid: chenodeoxycholic acid ratio, reduce the activation of Farnesoid X receptor (FXR) signaling in the liver, reduce insulin sensitivity, increase glycogen and lipogenesis, and reduce fatty acid oxidation in the liver [108]. At the same time, gut dysbiosis also inhibited FXR, reduced the secretion of fibroblast growth factor (FGF) 15/19, leading to fatty liver [109]. SCFAs, such as acetate, propionate and butyrate, are the products of bacterial fermentation of carbohydrate in the gut [110]. SCFAs in the intestine enter the liver through the portal vein, and acetate and propionate are precursors for fatty acid synthesis and gluconeogenesis, promoting liver fat accumulation [111]. Furthermore, SCFAs bind to G-protein coupled receptors of intestinal neuroendocrine L cells to secrete peptide YY and glucagon-like peptide 1(GLP-1), which promote nutrient absorption and liver fat generation [112]. Butyrate may activate the AMP-activated protein kinase (AMPK) pathway in the liver, leading to inhibition of oxidative stress and inflammation, upregulation of fatty acid oxidation, downregulation of fat synthesis genes, and reduced hepatosteatosis [113]. Interestingly, the patients with NAFLD and severe hepatic fibrosis had more acetate in the stool, while the patients with milder severity of NAFLD had
more SCFAs in butyrate and propionate [114]. Moreover, the concentration of ethanol in the blood of patients with NAFLD increased, and the bacteria *Proteobacteria*, which could produce ethanol, also tended to increase in the patients with steatohepatitis. Ethanol destroys the tight binding protein of the intestinal wall, increases the intestinal permeability, increases the endotoxin entering blood and the liver, leading to liver inflammation [115].

**Adipose tissue-liver axis**

Adipose tissues secrete adiponectin, leptin, and some pro-inflammatory cytokines such as interleukin (IL) 6 and TNF-α, which would influence the liver. Adiponectin binds to adiponectin receptor 1 and 2, respectively activates AMPK and PPAR-alpha pathways in the liver, and stimulate glucose use and fatty acid oxidation [116, 117]. Adiponectin also increases carnitine palmitoyltransferase I activity, enhances hepatic fatty acid oxidation, decreases the activities of acetyl-CoA carboxylase and fatty acid synthase [118]. However, adiponectin produced mainly from white adipose tissue is decreased in NASH patients [119]. When obesity develops, leptin secreted from white fatty tissue is increased to inhibit appetite and increase fatty acid oxidation [120]. However, in obese individuals, leptin resistance develops, and the increased leptin would exert proinflammatory activity. The serum leptin levels are positively associated with severity of liver inflammation and fibrosis [121, 122]. Leptin augments endothelin-1-induced contraction of HSCs [123]. Adipocytes also secrete TNF-α [124], which can increase insulin resistance and have pro-inflammatory effects [125]. TNF-α increased the gene expression of Mcp1, Tgfb1, and Timp1 in hepatocytes and the Tnf knockout improved glucose tolerance and significantly reduced the prevalence of hepatic steatosis and fibrosis in mice, indicating that TNF-α plays a role in the development and progression of NASH [126]. IL6 can be secreted from adipocytes and then increases macrophage infiltration of adipose tissue [127]. IL-6 infusion induces hepatic insulin resistance through increased adipose tissue lipolysis [128]. These data suggest that IL6 is
involved in the pathogenesis of hepatic insulin resistance.

**Renin-angiotensin system**

Hypertensive patients with biopsy-proven NAFLD on baseline RAS blockers had less advanced hepatic fibrosis [129]. Recently, a large retrospective study showed that angiotensin-converting enzyme inhibitors/angiotensin receptor blockers were associated with lower risk of hepatocellular carcinoma and cirrhotic complications in patients with NAFLD [130]. These data suggest a beneficial effect of RAS blockers in NAFLD. Transgenic hypertensive rats overexpressing the mouse renin gene with elevated levels of tissue angiotensin II developed hepatic steatosis, inflammation, and fibrosis [131]. The mice lacking the renin gene fed with high fat diet had decreased liver fat [132]. Aliskiren, a direct renin inhibitor, reduced hepatic steatosis in high-fat diet-fed mice and fibrosis in mice fed with methionine-choline-deficient diet [133, 134]. When renin or prorenin binds to the (pro)renin receptor (PRR), in addition to increasing the production and role of angiotensin (ANG II dependent pathway), it activates TGF-β, plasminogen activator inhibitor-1 (PAI-1), fibronectin, collagen I independently from Ang II (ANG II independent pathway) [135-137]. Our group found that PRR contributed to liver fibrosis and HSC activation, and its down-regulation attenuated liver fibrosis through inactivation of the ERK/TGF-β1/Smad3 pathway [138]. These results indicate that renin and prorenin can directly activate renin (pro) receptor-related intracellular signaling pathways, including ERK, TGF-β, cyclooxygenase2, fibronectin, collagen I, and PAI-1 independently of angiotensin II to induce fibrosis. Moreover, Ren et al. used N-acetylgalactosamine modified antisense oligonucleotides to suppress PRR expression in hepatocytes of high-fat diet-fed C57BL/6 mice and found that PRR inhibition reduced acetyl-CoA carboxylase and pyruvate desorption hydrogenase protein expression. This change reprogrammed liver lipid metabolism, resulting in reduced lipid synthesis and increased fatty acid oxidation. As a result, liver PRR suppression attenuated diet-induced obesity and fatty liver [139]. The proposed pathogenesis
involving from steatosis to fibrosis in patients with NAFLD is shown in Figure 1.

PROGRESSION OF NASH TO HCC

NASH is now the most common risk factor for HCC in USA [140]. The potential pathways linking NASH to HCC include chronic inflammation of the liver [141], alternations in immune response, lipid metabolism and gut microbiome [142], and genetic factor. Enhanced IL-6 and TNF production during NAFLD cause hepatic inflammation and activation of the oncogenic transcription factor STAT3 [143]. ER stress contributes to NASH-driven hepatic tumorigenesis via TNFR1 [144]. The hepatic oxidative DNA damage was increased in patients with NASH who developed HCC [145]. The unconventional prefoldin RPB5 interactor-induced DNA damage in hepatocytes triggered inflammation via T helper 17 lymphocytes and interleukin 17A, contributing to NASH and HCC development [146]. Furthermore, NAFLD caused a selective loss of intrahepatic CD4(+) but not CD8(+) T lymphocytes, which leaded to accelerated hepatocarcinogenesis [147]. Neutrophil infiltration was characterized in NASH-HCC and can exist in both tumor promoting and suppressing states [148]. Fatty acid accumulation increased junctional protein associated with coronary artery disease, leading to activation of Yes-associated protein 1 and tumor growth [149]. Dysregulated mammalian target of rapamycin (mTOR) stimulated sphingolipid and glycerophospholipid synthesis, leading to steatosis and HCC [150]. In NASH-driven HCC, metabolic reprogramming mediated by the downregulation of carnitine palmitoyltransferase 2 enables HCC cells to escape lipotoxicity and promotes hepatocarcinogenesis [151]. MicroRNA-21 can promote hepatic lipid accumulation and cancer progression by interacting with the sHbpl-p53-Srebp1c pathway [152]. The intestinal dysbiosis, gut permeability changes, and lipopolysaccharides translocation to the liver in NASH may increase secretion of the epiregulin growth factor, which triggered tumor hepatocyte proliferation [153]. Moreover, carriage of the PNPLA3 rs738409 C > G polymorphism is associated with greater
risk of NASH-HCC [154].

TREATMENT FOR NASH

Non-pharmacological therapy

Lifestyle modification

Lifestyle changes by eating less and exercise more to achieve weight loss remain the cornerstone of clinical care. Hypocaloric diet with a reduction of body weight decreased total body fat, visceral fat, and intrahepatic lipid content [155]. Some guidelines suggest restriction of energy by 1200–1500 kcal/day or a reduction of 500–1000 kcal/day to achieve weight loss [1, 156-158]. Weight reduction is beneficial for both non-obese (3-10%) [159] and obese patients (≥ 10%) [160, 161]. Other dietary compositions that may be beneficial for NAFLD includes omega-3 PUFA and coffee. Omega-3 PUFA has been shown to increase insulin sensitivity [162] and ameliorate steatohepatitis in experimental studies [163, 164]. One meta-analysis involving 9 studies with 355 patients showed decreased liver fat in patients with PUFA treatment [165]. Coffee is not only associated with a reduced risk of NAFLD but also decreased risk of liver fibrosis among patients with NAFLD [166, 167]. Regular exercise helps to enhance the effects of diet modifications. Physical activity with a target at least 150 min/week of moderate-intensity or 75–150 min/week of vigorous-intensity aerobic exercise is suggested [1, 156-158]. Both aerobic and resistance exercise reduce the hepatic fat content [168, 169]. In addition, the exercise intensity may be more important than duration or total volume [170]. In conclusion, lifestyle interventions to promote weight loss that include both diet and exercise is a proven therapeutic strategy to improve fatty liver disease.

Surgery

Bariatric surgery provides sustained and durable weight loss and improving obesity-related diseases [171, 172]. The most commonly performed bariatric procedures currently are
laparoscopic sleeve gastrectomy (LSG) and Roux-en-Y gastric bypass (RYGB). Two meta-analyses showed that bariatric surgery resulted in a biopsy-confirmed resolution of steatosis (56–66%), inflammation (45–50%), balloononing degeneration (49–76%), and fibrosis (25–40%) as well as reduction of nonalcoholic fatty liver disease activity (NAS) scores [173, 174]. A higher rate of improvement of steatosis and hepatic fibrosis was seen in Asian countries than non-Asian countries [174]. In addition, bariatric surgery was associated with decreased progression of NAFLD to cirrhosis [175] and reduced risks of any cancer and obesity-related cancer in NAFLD patients with severe obesity, particularly in cirrhotic patients [176]. However, new or worsening of NAFLD was seen in 12% of patients after bariatric surgery [173]. Bariatric surgery was associated with a significantly lower risk of incident major adverse liver outcomes (2.3% vs. 9.6% at 10 years) and major adverse cardiovascular events (8.5% vs. 15.7% at 10 years), as compared with nonsurgical management [177].

**Endoscopic therapy**

Endoscopic bariatric therapies, including intragastric balloons (IGB), endoscopic sleeve gastroplasty (ESG), duodenal mucosal resurfacing (DMR), and duodenal-Jejunal bypass liner (DJBL), were recently introduced as less invasive modalities to treat obesity and metabolic comorbidities. In a meta-analysis, improvement in steatosis and NAS were seen in 79.2% and 83.5% of patients receiving IGB, respectively [178]. Improvement of fibrosis for 1.5 stage by MR elastography was seen in 50% of patients with NAFLD after IGB placement [179]. ESG reduced body weight up to 15% and improved hepatic steatosis and fibrosis at two years of follow-up in obese patients with NAFLD [180]. Studies for efficacy and safety of ESG (NCT03426111; NCT04653311) and the comparison of ESG vs. LSG (NCT04060368) in patients with NASH are ongoing. DMR has been shown to reduce ALT, AST, and Fibrosis-4 scores in patients with diabetes mellitus [181]. Recently, an observational study enrolling 32 obese patients with diabetes mellitus who underwent DJBL showed improved non-invasive
markers of steatosis and NASH, but not fibrosis. The role of DJBL on NAFLD need further evaluation [182].

**Fecal microbiota transplantation**

Some studies have suggested that fecal transplantation help ameliorate steatohepatitis [183, 184]. A randomized controlled trial (RCT) using allogenic fecal microbiota transplantation (FMT) from lean vegan donors for patients with NAFLD through duodenal infusion found that there was no significant improvement in NAS, steatosis, and fibrosis scores. However, they observed a trend of improving necro-inflammatory scores and beneficial changes in hepatic gene expression and plasma metabolites involved in inflammation and lipid metabolism following allogenic FMT [185]. Another RCT using allogenic FMT via endoscopic duodenal infusion in patients with NAFLD found that FMT did not improve insulin resistance and hepatic steatosis but reduce small intestinal permeability at 6 months of follow-up [186].

**Pharmacological therapy**

The pharmacological agents predominantly target the following four mechanisms: (1) hepatic fat accumulation; (2) oxidative stress, inflammation, and apoptosis; (3) gut-liver axis, including bile acids, gut microbiomes and metabolic endotoxemia; and (4) hepatic fibrosis [187]. The agents targeting on different pathways are described below and those with promising results are summarized in Table 1.

**Agents targeting hepatic fat accumulation**

Pioglitazone, a PPAR γ agonist, improved hepatic steatosis, inflammation, and hepatocellular ballooning [188, 189]. Similar effects were found in Asian NASH patients [190]. In the phase 3 RESOLVE-IT trial, Elafibranor, a dual PPAR α/δ agonist, failed to
achieve NASH resolution [191]. Pemafibrate, a selective PPAR α modulator, did not decrease liver fat but caused a significant reduction in fibrosis for 6.2% [192]. Lanifibranor, a pan-PPAR agonist, significantly decrease the steatosis-activity-fibrosis activity (SAF-A) score for at least 2 points in 55% of the patients at 24 weeks [193].

GLP-1 agonists increase insulin secretion, inhibit glucagon secretion, delay gastric emptying, and decrease appetite. NASH resolution was seen in 39% of patients who received liraglutide for 48 weeks and in 59% of patients who received semaglutide for 72 weeks [194, 195]. However, fibrosis improvement was insignificant from both studies.

Sodium/glucose cotransporter 2 (SGLT2) inhibitors increase the urinary excretion of glucose. In a meta-analysis of ten RCTs, SGLT2 inhibitors can reduce aminotransferases and hepatic fat [196].

FGF19 and FGF21 are endocrines that regulate energy homeostasis. Aldafermin, a FGF19 analogue, led to reductions of liver fat content and a trend toward fibrosis improvement [197]. Pegbelfermin and efruxifermin are long acting, recombinant analogues of human FGF21 and both showed effects of reducing liver fat [198, 199].

Two phase IIa trials investigated the effects of acetyl-coenzyme A carboxylase (ACC) inhibitor monotherapy (PF-05221304) and combination with a diacylglycerol O-acyltransferase 2 (DGAT2) inhibitor (PF-06865571). Both PF-05221304 monotherapy or co-administration with PF-06865571 reduced liver fat content. [200]

Stearoyl-coenzyme A desaturase 1 (SCD-1) is a key enzyme that catalyzes the biosynthesis of monounsaturated fatty acids. A phase IIb trial (ARREST trial) showed that aramchol (a liver-targeted SCD-1 inhibitor) 600 mg did not cause significant reduction in liver fat content. Nevertheless, the observed change in liver histology and biochemical improvement suggests a potential role for aramchol in treating NASH and fibrosis [201].

Thyroid hormone receptor-β (THR-β) is predominantly expressed in the hepatocytes. Resmetirom, a selective THR-β agonist, significantly reduced more than 30% of hepatic fat
after 12 and 36 weeks of treatment in patients with NASH in phase II trial [202].

**Agents targeting oxidative stress, inflammation, and apoptosis**

Vitamin E, an antioxidative agent, had benefits on hepatic decompensation and transplant-free survival in patient with NASH [203]. In the PIVENS study which compared the effects of vitamin E, pioglitazone, and placebo in NASH patients without diabetes, showed that vitamin E (800 international units /day), but not pioglitazone, significantly improved nonalcoholic steatohepatitis [204].

Apoptosis signaling kinase 1 (ASK1) promote apoptosis, inflammation, and fibrosis in the liver. However, Selonsertib, an ASK1 inhibitor, failed to improve fibrosis in NASH patients with bridging fibrosis or compensated cirrhosis. [205]

Berberine ursodeoxycholate is an ionic salt of berberine and ursodeoxycholic acid. It reduced 4.8% of liver fat and improved glycemic control as well as liver enzymes in patients with NASH and diabetes [206].

**Agents targeting gut-liver axis**

In a phase IIb study, obeticholic acid (OCA), a FXR agonist, improved liver histology in 21% of NASH patients [207]. In patients with NASH and diabetes, OCA demonstrated the effects of increasing insulin sensitivity and reducing markers of liver inflammation as well as fibrosis [208]. In the interim analysis of a phase III trial, both 10mg and 25mg doses of OCA improved fibrosis (18% and 23%, respectively), but the NASH resolution endpoint was not met [209]. This study is ongoing to assess clinical outcomes.

**Agents targeting liver fibrosis**

Caspase is a protease that is associated with apoptosis and inflammation in the liver. However, emricasan, a pan-caspase inhibitor, did not improve fibrosis or resolution of NASH
Besides, for patients with NASH related cirrhosis and severe portal hypertension, emricasan did not improve hepatic venous pressure gradient (HVPG) or liver-related outcomes [211].

In a phase IIb CENTAUR trial, a 2-year study, cenicriviroc, a dual C-C chemokine receptor types 2 and 5 antagonist, achieved ≥ 1-stage of fibrosis improvement without worsening of NASH after one year of treatment compared with placebo (20% vs. 10%) [212]. Although fibrosis reduction was similar between cenicriviroc and placebo at the end of second year, twice the proportion on cenicriviroc who achieved fibrosis response at first year maintained fibrosis reduction at second year compared with placebo (60% vs. 30%, respectively) [213]. The long-term impact of cenicriviroc on fibrosis needs further evaluation.

Belapectin, a galectin-3 inhibitor, did not significantly reduce HVPG or fibrosis in patients with NASH, cirrhosis and portal hypertension; however, in a subgroup of patients without esophageal varices, belapectin reduced HVPG and esophageal varices development [214].

Information about the ongoing phase III clinical trials of promising drugs on phase II studies are listed in Table 2.

Combination therapy

NAFLD is a multifactorial disease and combining therapies with different targets may have additional benefits than monotherapies [215]. Cilofexor (FXR agonist) plus firsocostat (ACC inhibitor) led to improvements in NASH activity compared to placebo, or single agent in patients with bridging fibrosis and cirrhosis [216]. Semaglutide with firsocostat and/or cilofexor showed greater improvements in liver steatosis, liver biochemistry than semaglutide alone [217]. Combined ACC inhibitors and DGA T2 inhibitors reduced liver fat content and mitigated the side effect of elevated serum triglycerides [200].
PERSPECTIVES

As understanding of mechanisms of NASH and its fibrosis increases, more therapies will be introduced and tested in clinical trials. The pathogenesis of NASH and fibrosis is complex; therefore, it would be difficult to treat the disease through one therapy. Combination therapy is the focus of future development of treatment. Furthermore, better care of extra-hepatic complications of NASH, novel biomarkers for diagnosis, risk stratification and treatment responses, and more clinical trials in Asian groups also need to be well researched and developed.

AUTHORS’ CONTRIBUTION

KC Lee and PS Wu drafted the manuscript and HC Lin revised the manuscript. All the authors read and approved the final version.
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Bariatric Surgery With Major Adverse Liver and Cardiovascular Outcomes in Patients With

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insulin resistance and estimated hepatic steatosis and fibrosis after endoscopic sleeve

Endoscopic duodenal mucosal resurfacing improves glycaemic and hepatic indices in type 2


Figure 1. The progression of hepatic steatosis to inflammation and fibrosis in liver. Both metabolic and genetic factors contribute to the formation of hepatic steatosis. Fat accumulation in hepatocytes leads to organelles dysfunction and lipotoxicity. Then oxidative stress species or signaling molecules transmitted through extracellular vesicles or diffusion, activating other parenchymal and non-parenchymal cells, which subsequently causes inflammatory cascades, steatohepatitis and liver fibrosis. On the other hand, gut-derived bacterial end-products, metabolites, gut hormones, adipose tissue-derived cytokines or adipokines and renin-angiotensin-system all contribute the progression from steatosis to inflammation and fibrosis. SFA: saturated fatty acid; TG: triglyceride; ER: endoplasmic reticulum; HH-OPN: Hedgehog-osteopontin; KC: Kupffer cell; HSC: hepatic stellate cell.
<table>
<thead>
<tr>
<th>Type of drug</th>
<th>Mechanism of action</th>
<th>Drug name</th>
<th>Study design</th>
<th>Study outcome</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>PPAR agonist</strong></td>
<td>PPAR-γ: ↑insulin sensitivity, modulate adipose tissue distribution, PPAR-α: ↑fatty acid β-oxidation, PPAR-δ: anti-inflammatory</td>
<td>Pioglitazone (PPAR-γ agonist)</td>
<td>RCT; NASH, prediabetes/DM</td>
<td>↓ALT/AST</td>
<td>[188]</td>
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<td></td>
<td></td>
<td>Pioglitazone vs. placebo</td>
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<td>↓Steatosis, ballooning necrosis and inflammation Fibrosis not improved</td>
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<td></td>
<td></td>
<td>Pioglitazone vs. placebo</td>
<td></td>
<td>↓ALT/GGT</td>
<td>[189]</td>
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<td></td>
<td></td>
<td>Pemafibrate (SPPARMα)</td>
<td>Phase II RCT; NAFLD and ↑ALT</td>
<td>↓ALT</td>
<td>[192]</td>
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<tr>
<td></td>
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<td>Pemafibrate vs. placebo</td>
<td></td>
<td>↓Liver stiffness</td>
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<td></td>
<td>Lanifibranor (Pan-PPAR agonist)</td>
<td>Phase IIb RCT; NASH (SAF-A ≥ 3) Lanifibranor vs. placebo</td>
<td>↓SAF-A score ≥ 2 points in lanifibranor 1200mg group</td>
<td>[193]</td>
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<tr>
<td><strong>GLP-1 agonist</strong></td>
<td>↑Insulin secretion, ↓Glucagon secretion, ↓Gastric emptying, ↓Appetite</td>
<td>Liraglutide</td>
<td>Phase II RCT; NASH, obesity Liraglutide vs. placebo</td>
<td>↑Resolution of NASH without worsening of fibrosis</td>
<td>[194]</td>
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<td></td>
<td></td>
<td>Liraglutide vs. placebo</td>
<td></td>
<td>No difference in fibrosis improvement</td>
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<td>Semaglutide</td>
<td>RCT, phase II; NASH (F1-F3 fibrosis) Semaglutide vs. placebo</td>
<td>↑Resolution of NASH without worsening of fibrosis in semaglutide 0.4mg group</td>
<td>[195]</td>
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<td>No difference in fibrosis improvement</td>
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<tr>
<td>Treatment Type</td>
<td>Functional Changes</td>
<td>Baseline Study Details</td>
<td>Outcome Measures</td>
<td>References</td>
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<tr>
<td><strong>SGLT-2 inhibitor</strong></td>
<td>↑ Urinary excretion of glucose</td>
<td>SGLT-2 inhibitors Meta-analysis of 10 RCTs; NAFLD, DM SGLT-2 inhibitor vs. other antidiabetic drugs</td>
<td>↓ALT/AST, ↓Liver fat content, visceral fat, and subcutaneous fat areas</td>
<td>[196]</td>
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<tr>
<td><strong>FGF-19 analogue</strong></td>
<td>↓ Bile acids synthesis, ↓ Hepatic gluconeogenesis, ↑ Fatty acid oxidation</td>
<td>Aldafermin Phase II RCT; NASH (NAS ≥ 4, F2-F3 fibrosis, and liver fat content ≥ 8%) Aldafermin vs. placebo</td>
<td>↓ALT/AST, ↓Liver fat fraction on MRI-PDFF</td>
<td>[197]</td>
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<td><strong>FGF-21 analogue</strong></td>
<td>↓ Hepatic gluconeogenesis, ↑ Insulin sensitivity, ↑ Energy expenditure, ↑ Mitochondria beta-oxidation in hepatocytes</td>
<td>Pegbelfermin Phase IIa RCT; NASH (F1-3 fibrosis, and liver fat content ≥ 10%), obesity Pegbelfermin vs. placebo</td>
<td>↓Liver fat fraction on MRI-PDFF</td>
<td>[198]</td>
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<tr>
<td><strong>Acetyl-CoA carboxylase inhibitor</strong></td>
<td>↓DNL, ↑ Fatty acid oxidation</td>
<td>PF-05221304 2 phase IIa RCTs; NAFLD/NASH PF-05221304 monotherapy vs. placebo</td>
<td>PF-05221304 monotherapy: ↓Liver fat on MRI-PDFF but ↑TG</td>
<td>[200]</td>
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<tr>
<td><strong>Diacylglycerol acyltransferase 2 inhibitor</strong></td>
<td>↓Synthesis of fatty acids into TGs</td>
<td>PF-06865571 PF-05221304 and PF-06865571 co-administration vs. placebo</td>
<td>Co-administration therapy: ↓Liver fat and mitigated ACC inhibitor-mediated effect on TG</td>
<td>[201]</td>
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<td><strong>Stearoyl-CoA desaturase 1 inhibitor</strong></td>
<td>↓DNL</td>
<td>Aramchol Phase IIb RCT; NASH Aramchol vs. placebo</td>
<td>↓Liver fat content in aramchol 600mg group but not significant.</td>
<td>[201]</td>
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<tr>
<td><strong>Selective thyroid hormone receptor-β</strong></td>
<td>↓ LDL, cholesterol, and TG, ↑ Fatty acid oxidation</td>
<td>Resmetirom Phase II RCT; NASH (F1-F3 fibrosis, and liver fat content ≥ 10%)</td>
<td>↓Liver fat on MRI-PDFF</td>
<td>[202]</td>
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<tr>
<td>agonist</td>
<td>Resmetirom vs. placebo</td>
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<td><strong>Anti-oxidant</strong></td>
<td>Vitamin E</td>
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<td>Anti-oxidative stress</td>
<td>RCT; NASH, no DM</td>
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<td>Vitamin E vs. pioglitazone vs. placebo</td>
<td>↓ALT/AST for both vitamin E and pioglitazone groups</td>
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<td></td>
<td>NASH improvement in vitamin E group but not in pioglitazone group</td>
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<td></td>
<td>Fibrosis not improved in vitamin E and pioglitazone groups</td>
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<td><strong>Bile acid analogue</strong></td>
<td>Berberine ursodeoxycholate</td>
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<tr>
<td>Anti-inflammation</td>
<td>Phase II RCT; NAFLD, DM</td>
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<tr>
<td>Berberine ursodeoxycholate vs. placebo</td>
<td>↓Liver fat content on MRI-PDFF [206]</td>
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<tr>
<td></td>
<td>↓DNL</td>
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<td></td>
<td>↑Fatty acid β-oxidation</td>
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<td></td>
<td>↑Cholesterol excretion</td>
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<td>Obeticholic acid (FXR agonist)</td>
<td>Phase III RCT; NASH (NAS ≥ 4, F2-F3 fibrosis or F1 with ≥ 1 accompanying comorbidity)</td>
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<tr>
<td>Obeticholic acid vs. placebo</td>
<td>↓Fibrosis</td>
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<tr>
<td></td>
<td>No significant resolution of NASH</td>
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</table>

NAFLD, non-alcoholic fatty liver disease; NASH, nonalcoholic steatohepatitis; PPAR, peroxisome proliferator-activated receptor; RCT, randomized controlled trial; DM, diabetes mellitus; ALT, alanine transaminase; AST, aspartate aminotransferase; GGT, gamma glutamyl transpeptidase; SPPARMα, selective peroxisome proliferator-activated receptor alpha modulator; NAS, nonalcoholic fatty liver disease activity score; SAF-A score, steatosis-activity-fibrosis activity score; GLP-1, glucagon-like peptide-1; SGLT-2, sodium–glucose cotransporter 2 inhibitors; FGF, fibroblast growth factor; MRI-PDFF, magnetic resonance imaging proton density fat fraction; DGAT2, diacylglycerol acyltransferase 2; DNL, de novo lipogenesis; TG, triglyceride; FXR, farnesoid X receptor
Table 2. Ongoing phase III clinical trials of pharmacological agents in patients with NAFLD or NASH

| Agent          | Mechanism                  | Patient                                           | Outcome                                                               | Status   | ClinicalTrials.gov Identifier |
|----------------|----------------------------|---------------------------------------------------|                                                                      |          |                              |
| Lanifibranor   | Pan-PPAR agonist           | NASH with stage 2-3 fibrosis without cirrhosis    | NASH resolution; Fibrosis improvement; liver-related events          | Recruiting | NCT04849728                   |
|                |                            |                                                   |                                                                      |          |                              |
| Semaglutide    | GLP-1 agonist              | NASH with stage 2-3 fibrosis                     | NASH resolution; Fibrosis improvement; liver-related events          | Recruiting | NCT04822181                   |
|                |                            |                                                   |                                                                      |          |                              |
| Dapagliflozin  | SGLT-2 inhibitor           | NASH and type 2 DM, without cirrhosis             | Histology improvement                                               | Recruiting | NCT03723252                   |
|                |                            |                                                   |                                                                      |          |                              |
| Resmetirom     | Selective thyroid hormone receptor-β agonist | NASH without cirrhosis                          | NASH resolution; liver-related events                                | Recruiting | NCT03900429                   |
| Aramchol       | Stearoyl-CoA desaturase 1 inhibitor | NASH with stage 2-3 fibrosis without cirrhosis; type 2 DM or prediabetes | NASH resolution; Fibrosis improvement; liver-related events          | Recruiting | NCT04104321                   |
| Belapectin     | Galectin-3 inhibitor       | NASH cirrhosis without esophageal or gastric varices | Newly developed esophageal varices                                   | Recruiting | NCT04365868                   |
| Oltipraz       | Liver X receptor alpha-inhibitor | NAFLD without cirrhosis                          | Liver fat                                                            | Recruiting | NCT04142749                   |

NAFLD, nonalcoholic fatty liver disease; NASH, nonalcoholic steatohepatitis; SGLT-2, sodium–glucose cotransporter 2 inhibitors; DM, diabetes mellitus; GLP-1, glucagon-like peptide-1; PPAR, peroxisome proliferator-activated receptor