Updates in the quantitative assessment of liver fibrosis for nonalcoholic fatty liver disease: Histological perspective

Gwyneth Soon1 and Aileen Wee1,2

1Department of Pathology, National University Hospital, Singapore; 2Department of Pathology, Yong Loo Lin School of Medicine, National University of Singapore, Singapore, Singapore

Nonalcoholic fatty liver disease/nonalcoholic steatohepatitis (NAFLD/NASH) is a major cause of liver fibrosis and cirrhosis. Accurate assessment of liver fibrosis is important for predicting disease outcomes and assessing therapeutic response in clinical practice and clinical trials. Although noninvasive tests such as transient elastography and magnetic resonance elastography are preferred where possible, histological assessment of liver fibrosis via semiquantitative scoring systems remains the current gold standard. Collagen proportionate area provides more granularity by measuring the percentage of fibrosis on a continuous scale, but is limited by the absence of architectural input. Although not yet used in routine clinical practice, advances in second harmonic generation/two-photon excitation fluorescence (SHG/TPEF) microscopy imaging show great promise in characterising architectural features of fibrosis at the individual collagen fiber level. Quantification and calculation of different detailed variables of collagen fibers can be used to establish algorithm-based quantitative fibrosis scores (e.g., qFibrosis, q-FPs), which have been validated against fibrosis stage in NAFLD. Artificial intelligence is being explored to further refine and develop quantitative fibrosis scoring methods. SHG-microscopy shows promise as the new gold standard for the quantitative measurement of liver fibrosis. This has reaffirmed the pivotal role of the liver biopsy in fibrosis assessment in NAFLD, at least for the near-future. The ability of SHG-derived algorithms to intuitively detect subtle nuances in liver fibrosis changes over a continuous scale should be employed to redress the efficacy endpoint for fibrosis in NASH clinical trials; this approach may improve the outcomes of the trials evaluating therapeutic response to antifibrotic drugs. (Clin Mol Hepatol 2021;27:44-57)

Keywords: Nonalcoholic fatty liver disease; Fibrosis; Biopsy

Abbreviations:
ALI, artificial intelligence; ASMA, antibodies against α-smooth muscle cells; CARS, coherent anti-Stokes Raman scattering; CHB, chronic Hepatitis B; CHC, chronic hepatitis C; CPA, collagen proportionate area; ECM, extracellular matrix; ELF, enhanced liver fibrosis; FIB-4, fibrosis-4; FLIP, Fatty Liver: Inhibition of Progression; FSR, fractional synthesis rate; HSC, hepatic stellate cells; LSEC, liver sinusoidal endothelial cells; MRE, magnetic resonance elastography; NAFLD, nonalcoholic fatty liver disease; NAS, nonalcoholic fatty liver disease activity score; NASH CRN, Nonalcoholic Steatohepatitis Clinical Research Network; NASH, nonalcoholic steatohepatitis; NFS, nonalcoholic fatty liver disease fibrosis score; NIDDK, National Institute of Diabetes and Digestive and Kidney Diseases; q-FPs, quantification of fibrosis-related parameters; SAF, steatosis activity fibrosis; SHG, second harmonic generation; TE, transient elastography; TPEF, two-photon excitation fluorescence

Corresponding author: Aileen Wee
Department of Pathology, National University Hospital, 5 Lower Kent Ridge Road, Singapore 119074, Singapore
Tel: +65-67724305, Fax: +65-67780671
E-mail: aileen_wee@nuhs.edu.sg
https://orcid.org/0000-0003-3107-7599

Editor: Han Ah Lee, Korea University College of Medicine, Korea

Received: Jul. 20, 2020 / Revised: Aug. 25, 2020 / Accepted: Aug. 26, 2020

Copyright © 2021 by Korean Association for the Study of the Liver
This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/3.0/) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.
INTRODUCTION

Nonalcoholic fatty liver disease (NAFLD) is a burgeoning global health concern in tandem with metabolic syndrome.¹ NAFLD encompasses steatosis to nonalcoholic steatohepatitis (NASH); the latter may progress to cirrhosis. In the United States, NAFLD is one of the leading indications for liver transplantation.² Prevalence of NAFLD in Asia has increased from 25% (1999–2005) to 34% (2012–2017).³ The histological diagnosis of NASH is based on four key parameters, viz., steatosis, hepatocellular ballooning, lobular inflammation, and fibrosis. Precise grading of severity and fibrosis staging are crucial for understanding the pathogenesis and development of therapeutic targets to halt, reverse as well as prevent disease.

Accurate quantitative assessment of liver fibrosis in NAFLD is important to predict risk of developing complications and track disease progression, especially as an efficacy endpoint in clinical trials of antifibrotic agents. Fibrosis stage is the most important histological predictor of liver-specific outcome; the risk of cirrhotic decompensation and hepatocellular carcinoma increases with fibrosis stage.⁴⁷

Noninvasive methods of fibrosis assessment are recommended by international management guidelines for NAFLD,⁸⁻¹² given that only a minority of patients would eventually develop liver-related complications, and progression to cirrhosis takes decades.¹³⁻¹⁶ However, it is increasingly recognised that the liver biopsy, despite its limitations, not only remains the ‘gold standard’ but provides a window into the architectural pattern and nature of fibrosis far beyond the scope of current routine histopathology practice, particularly with recent advances in techniques such as second harmonic generation (SHG)-microscopy and advanced computing systems. Bedossa has reiterated the importance of liver biopsy in the diagnosis and management of NAFLD/NASH in (selected) patients and clinical trials.¹⁷ Wanless in a recent editorial also stated categorically that “quantitative biopsy assessment using SHG-microscopy should be considered the new gold standard for the measurement of liver fibrosis”.²⁸

There are many unmet needs concerning various aspects of NASH.¹⁹ The current U.S. Food and Drug Administration requirements regarding efficacy endpoints to demonstrate therapeutic response in NASH clinical trials may have to be re-addressed in favour of more precise and granular criteria on a continuous scale with the aid of advancing laser microscopy and machine learning. This review aims to focus on current and developing techniques that assess fibrosis on liver tissue samples.

PATHOGENESIS OF LIVER FIBROSIS IN NAFLD

Hepatocellular injury and inflammation are supposedly the primary drivers of fibrogenesis in NASH patients, finally culminating in cirrhosis. The PIVENS and FLINT clinical trials have demonstrated the strong link between histological resolution of steatohepatitis (NAFLD activity score ≥2) with at least 1-stage decrease or more in fibrosis in NASH.²⁰ Liver fibrosis is a dynamic wound healing response to chronic liver injury characterized by accumulation of extracellular matrix (ECM) components, including collagen fibers, sulfated proteoglycans and glycoproteins, in excess of matrix degradation by enzymes. Activation of hepatic stellate cells (HSC) during liver injury and inflammation from their quiescent state to proliferative, fibrogenic and contractile myofibroblasts, in conjunction with endothelial cells, Kupffer cell infiltration and activation, and secretion of other inflammatory molecules, is the key event in fibrogenesis.²¹⁻²²

The adult NAFLD pattern of fibrosis starts as delicate perisinusoidal deposition of collagen fibers in the space of Disse in the centrilobular zone (zone 3) of the hepatic lobule.²³ Periportal fibrosis then develops with short collagen strands extending into zone 1, and thereafter, bridging with central veins via fibrous septa. Cirrhosis eventually ensues in a small percentage of cases. In comparison, the fibrosis is zone 1-centric in (pre-adolescent) children, beginning around portal regions and progressing to bridging fibrosis.

NONINVASIVE TESTS FOR FIBROSIS ASSESSMENT

Imaging modalities often use liver stiffness as a surrogate measure to assess the amount of liver fibrosis.²⁴ Vibration-controlled transient elastography (TE) ( FibroScan®) is one of the most commonly used techniques.²⁵ Magnetic resonance elastography (MRE) has been shown to be superior to TE in that it can diagnose earlier stages of fibrosis, evaluate a larger portion of the liver, and provide for fat quantification via MR spectroscopy.²⁶⁻²⁸ Other newer techniques include multiparametric magnetic resonance imaging.²⁹⁻卅 Composite scoring systems based on serum biomarkers, such as fibrosis-4 (FIB-4) index, NAFLD fibrosis score (NFS), and enhanced liver fibrosis (ELF) test, have also been developed as surrogate measures of fibrosis severity in NAFLD.³¹⁻³⁶ Recently, there has been interest in gut microbiome to identify signatures for advanced fibrosis in NAFLD,³⁷⁻³⁸ as well as proteomic or multi-
omic studies.\textsuperscript{39,40} However, these methods are all indirect measures of fibrosis encumbered by confounders and lack of specificity.

Some of the most pressing issues with noninvasive tests are difficulty in determining the optimum cut-off to differentiate intermediate stages of fibrosis; inability to reflect architectural changes/fibrosis stage that may not correspond with amount of collagen deposition; and the incapability to assess the dynamic nature of fibrosis progression or regression. These data are essential in tracking disease progression, prognostication, identifying patients who are most likely to benefit from any therapeutic intervention, and assessing the efficacy of antifibrotic drugs.\textsuperscript{13,41,42} Recent advances in molecular and targeted MR imaging studies of small molecules that bind to fibrillary collagen or other ECM proteins are promising noninvasive approaches in the direct detection and quantification of liver fibrosis that would help overcome some of these limitations.\textsuperscript{43-45}

HISTOLOGICAL SCORING SYSTEMS BASED ON LIGHT MICROSCOPY

The performance of noninvasive tests is still currently measured against the ‘gold standard’ – histological assessment of liver tissue. The existing histological scoring systems for assessing fibrosis in NAFLD are semiquantitative categorical systems that have been developed to standardize and improve observer variability, especially in clinical trials (Table 1). Masson Trichrome or other connective tissue stains are performed to highlight fibrosis, particularly the delicate perisinusoidal component.

Strictly speaking, as categorical labels, histological scoring systems do not quantify the absolute amount of fibrosis across the entire liver, but are rather a reflection of the architectural patterns in the scarred liver. Their accuracy therefore relies not only on a pathologist’s judgment but also on sample adequacy. Besides the sampling variability inherent in a biopsy procedure and disease heterogeneity,\textsuperscript{46,47} suboptimal samples exacerbate the problem of inter- and intra-observer variability.\textsuperscript{48} Both the length and diameter of the cores must be sufficient for adequate assessment of portal tracts and central veins. A core length of 25 mm obtained via a 16-gauge needle is most ideal.\textsuperscript{49-52}

Brunt system

The Brunt system was the first histological assessment system proposed to categorize the morphologic features of NASH for grading and staging the disease.\textsuperscript{53} It was intended to be used only after overall evaluation of the biopsy had established a diagnosis of NASH. Perisinusoidal/pericellular fibrosis was recognised as the earliest stage of fibrosis, with subsequent progression to periportal fibrosis, bridging fibrosis, and cirrhosis.

Nonalcoholic Steatohepatitis Clinical Research Network (NASH CRN): NAFLD activity score (NAS)

The Brunt system was further refined to include the full spectrum of NAFLD and be applicable to adults and children.\textsuperscript{48} This new scoring system, developed and validated by the National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK) sponsored NASH CRN Pathology Committee, is currently the most recognised system for scoring NAFLD in clinical trials and experimental studies.\textsuperscript{23,54} The score is similarly not meant to supplant the pathologist’s diagnosis of NASH.\textsuperscript{55} The NAS is a summative score of three components: steatosis, ballooning and lobular inflammation. Fibrosis is described separately using a 5-stage system (0 to 4), with further subclassification of stage 1 into substages a–c depending on the location of collagen deposition.

<table>
<thead>
<tr>
<th>Fibrosis stage</th>
<th>Brunt system\textsuperscript{53}</th>
<th>NASH CRN score\textsuperscript{48}</th>
<th>SAF score\textsuperscript{57}</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>None</td>
<td>None</td>
<td>None</td>
</tr>
<tr>
<td>1</td>
<td>Zone 3 perisinusoidal fibrosis; focally or extensively present</td>
<td>1a: Mild (delicate) zone 3 perisinusoidal fibrosis</td>
<td>1c: Portal fibrosis only</td>
</tr>
<tr>
<td></td>
<td></td>
<td>1b: Moderate (dense) zone 3 perisinusoidal fibrosis</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>Zone 3 perisinusoidal fibrosis with focal or extensive periportal fibrosis</td>
<td>Zone 3 perisinusoidal fibrosis with periportal fibrosis</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>Zone 3 perisinusoidal fibrosis and portal fibrosis with focal or extensive bridging fibrosis</td>
<td>Bridging fibrosis</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>Cirrhosis</td>
<td>Cirrhosis</td>
<td></td>
</tr>
</tbody>
</table>

NASH CRN, Nonalcoholic Steatohepatitis Clinical Research Network; SAF, steatosis activity fibrosis.
The Fatty Liver: Inhibition of Progression (FLIP) consortium: steatosis activity fibrosis (SAF) score

The SAF scoring system, developed by the FLIP European consortium, removes steatosis from the activity score of NAS. It proposes an algorithm to distinguish patients with or without NASH, based on the activity alone (ballooning and lobular inflammation); the three individually scored components of steatosis, activity and fibrosis then comprise the SAF score. The fibrosis component is staged similarly as in the NASH CRN system; the inclusion of fibrosis in the SAF score presents a snapshot of the patient’s disease status. This scoring system was recently clinically validated in patients enrolled in a therapeutic trial.

Limitations of histological assessment scoring systems

Fibrosis deposition is actually a continuum that may not fall neatly into the designated categories of these semiquantitative staging systems. Distinguishing between the intermediate stages of fibrosis is challenging on liver biopsies. The single numeric value of the staging system does not convey the variability and extent of changes seen within each stage, nor the architectural/vascular alterations present. Moreover, the degree of perisinusoidal fibrosis is not captured in the higher stages of fibrosis. The severity of perisinusoidal fibrosis may contribute to portal hypertension in the absence of advanced fibrosis. Measurement of perisinusoidal fibrosis at all stages might enhance the evaluation of fibrosis outcome in clinical trial endpoints.

Fibrosis is a dynamic process with both progression and regression occurring concurrently, together with parenchymal remodeling, even at the ‘final’ cirrhotic stage. In this respect, Sun et al. proposed the Beijing classification based on chronic hepatitis B (CHB) patients to reflect the quality of fibrosis in advanced stage disease, in order to predict patient outcomes. However, this system has yet to be validated in NAFLD; furthermore, it is only applicable for later stage disease after parenchymal extinction and its sequelae have occurred.

Fibrosis is also a relatively slow process; current scoring systems may not provide sufficient granularity for assessing subtle changes during follow-up studies. Furthermore, in current practice, the amount of fibrosis assessed based on the few 3 to 4-microns thick sections of the biopsy core examined is assumed to be representative of the entire core, and by extension, the rest of the patient’s liver; truly a ‘sample’ of a ‘sample’. Lastly, traditional light microcopy simply displays the presence of collagen fibers via connective tissue stains; it does not unveil collagen fiber structure nor the status of other cellular players in this fibrogenesis process.

MORPHOMETRIC ANALYSIS

Collagen proportionate area (CPA) measurement is the most validated morphometric approach that quantifies the amount of fibrous tissue present as a proportion of the total biopsy area, after subtracting for structural collagen (e.g., in large vessel walls). This automated process employs digital image analysis on histological sections, most commonly stained with Picro-Sirius red to highlight collagen. CPA has been validated against hepatic venous pressure gradient measurements and clinical outcomes, mainly in patients with chronic hepatitis C (CHC). More recently, Buzzetti et al. demonstrated CPA as an independent predictor of long-term outcome in NAFLD, including early stage disease.

CPA provides a linear quantification of fibrosis that has the sensitivity to detect small variations in the amount of collagen and fibrosis, which is especially useful in clinical trials that are often of relatively short duration. Furthermore, some clinical trials currently use a reduction of fibrosis histological stage by 1 or more as an outcome; however, a reduction in stage from 4 to 3 may have different implications regarding the efficacy of an antifibrotic drug as compared to a reduction from stage 2 to 1, given that CPA increases exponentially by fibrosis stage. As a caveat though, sample adequacy becomes important when dealing with such precise measurements; moreover, adequate sample size differs between etiologies of cirrhosis. CPA measurement is also still subject to technical issues such as variances in staining procedure, operator experience, and imaging software used.

CPA is unable to evaluate architectural changes such as bridging fibrosis and nodularity, Masugi et al. demonstrated a strong but nonlinear relationship between Brunt fibrosis stage and the combined area ratio of collagen and elastin fibers, measured on Elastica van Gieson-stained biopsies from NAFLD patients. There was a marked difference between stages 4 and 3, but much smaller differences among stages 0–3. This was attributed to the relatively limited areas of bridging fibrosis in stage 3; thus, despite the altered architecture, the absolute amount of fibrosis may not differ significantly from earlier stages.

Hence, while CPA is a sensitive linear measurement that directly quantifies fibrosis, it does not provide any information on spatial alterations, the dynamic nature of fibrosis or the other cellular...
components involved in this process. It is still subject to the same potential sampling error that traditional histology suffers from.

**MULTIPLE LASER-BASED MICROSCOPY**

**SHG/two-photon excitation fluorescence (TPEF) laser microscopy**

Recent advancement in ultra-fast lasers have enabled imaging of optical signals, such as autofluorescence, from unstained, paraffin-embedded histological sections of biological samples. The ability to characterize the unique crystalline triple-helix structure and intrinsic optical signals of fibrillar collagen have opened up a whole new realm for quantitative assessment of liver fibrosis, re-establishing the importance of the liver biopsy. SHG microscopy is a nonlinear optical tissue imaging system that enables automated quantification of fibrosis based on the unique architectural features of collagen. TPEF permits visualization of the background liver architecture through endogenous tissue signals. SHG/TPEF microscopy therefore allows identification of individual collagen fibers, localization of collagen in 2D and 3D formats, and quantification of their physical attributes, such as number, length, diameter, orientation, contour, and cross-linkages of the collagen fibers with each other. Characterization of the collagen fibers and relationship of aggregated thick and dispersed thin fibers within a 3D lattice framework provides insights into the remodeling dynamism and would be of great pathomechanistic and pharmaceutical value.

There has been a surge in studies utilising SHG to assess liver fibrosis in the past decade (Fig. 1, Table 2). One of the most pivotal developments was the establishment of “qFibrosis” by Xu et al. - a fully-quantitative stain-free method for the automated assessment of liver fibrosis that incorporates spatial architectural features of pathological relevance at the tissue level. In order to achieve this concept of histological staging, the qFibrosis design utilised SHG/TPEF for 1) detection of changes in the respective collagen patterns and 2) quantitative identification of histopathological architectural features. Eighty-seven collagen architectural features were selected and categorized into three groups, namely, portal, septal, and fibrillar collagen, and these features are translated into quantitative parameters to build up the three respective subindices which were then combined into a single index, qFibrosis.

qFibrosis, first validated on core biopsies from CHB patients, was shown to reliably stage liver fibrosis with reduced variability of sampling error and inter-/intra-observer bias, as well as differentiate intra-stage cirrhosis changes, an essential step for monitoring progression or regression of cirrhosis and response to antifibrotic drugs. The successful demonstration of qFibrosis as a reproducible and robust tool, outperforming CPA and (hepato)pathologists’ observer variability in CHB and CHC, laid the groundwork for SHG microscopy-based assessment of liver fibrosis in NAFLD.

Pirhonen et al. subsequently performed a proof-of-concept study in NAFLD patients using SHG and coherent anti-Stokes Raman scattering (CARS) microscopy. They focused on patients with no or early stage fibrosis, as early detection of fibrosis is crucial for identifying individuals at risk for advanced liver disease. SHG and CARS imaging could detect fibrillar collagen and fat, respectively, in a label-free manner, allowing for automated and sensitive quantification of early fibrosis with continuous grading in NAFLD.

In 2017, Wang et al. then proposed a validated SHG-based quantification of fibrosis-related parameters (q-FPs) model in NAFLD that offered a more refined assessment of collagen architectural changes along a continuous, quantitative scale. The authors showed that SHG could differentiate subtle differences between fibrosis stages 1a, 1b, and 1c (NASH CRN system) and differences in zonal distribution of fibrosis in patients with cirrhosis. In a 2020 validation study, Wang et al. further showed that q-FF was highly accurate in assessing different stages of fibrosis in NAFLD patients and correlated strongly with histological scoring and liver stiffness measurement.

Similarly, Chang et al. developed a fibrosis index (SHG B-index) comprising 14 unique SHG-based collagen parameters that correlated with severity of NAFLD fibrosis in a continuous fashion, validating the reliability of SHG as an automated method for fibrosis staging. In their cross-validation analysis, the SHG B-index demonstrated high specificity for diagnosis of all grades of fibrosis (Brunt fibrosis stage), although it was less discerning in discriminating between early stages of fibrosis. The advantage of the SHG B-index is that it provides a convenient single composite index that can be easily applied both in routine practice and clinical trials.

Well-defined, accurate, reproducible, and clinically meaningful endpoints are currently lacking to assess efficacy of NASH clinical trials. Besides fibrosis, histological assessment of NASH severity is used as endpoints in clinical trials, despite acknowledged issues with accuracy and reproducibility of these parameters amongst
Figure 1. Comparison of histopathological staining (H&E and Masson trichrome) with SHG/TPEF images of liver biopsy tissue from NAFLD patients. SHG imaging, Masson trichrome, and H&E were performed on serial sections (×200). H&E, haemotoxylin and eosin; SHG, second harmonic generation; TPEF, two-photon excitation fluorescence; NAFLD, nonalcoholic fatty liver disease.
Table 2. Performance of SHG-based models for quantitative assessment of liver fibrosis in NAFLD

<table>
<thead>
<tr>
<th>Study</th>
<th>Model</th>
<th>Methodology</th>
<th>No. of patients</th>
<th>Performance</th>
</tr>
</thead>
<tbody>
<tr>
<td>Wang et al. (2017)</td>
<td>q-FP</td>
<td>SHG/TPF to capture images (of whole biopsy section)</td>
<td>50 (test cohort)</td>
<td>Principal component analysis model of 16 q-FPs:</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Images assessed with computerized image-analysis by two independent investigators to output the profile</td>
<td>42 (validation cohort)</td>
<td>- Fibrosis vs. no fibrosis: AUC 0.88</td>
</tr>
<tr>
<td></td>
<td></td>
<td>of q-FPs data for each slice in operator-defined segmentation regions of liver tissue, including:</td>
<td></td>
<td>- Cirrhosis vs. earlier stages: AUC 0.93</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(1) General: the liver section in its entirety</td>
<td></td>
<td>Linear scale of fibrosis measurement of 4 q-FPs using desirability functions:</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(2) Perisinusoidal: hepatocyte-associated collagen in the perisinusoidal space</td>
<td></td>
<td>- Related to fibrosis stage (P&lt;0.0001)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(3) Vessel: collagen fibrils directly connected to veins; and</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>(4) Vessel bridges: collagen fibrils extending from vein to vein or vein to portal tract.</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>70 q-FPs had interclass concordance ≥0.8 which were selected for further model development</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>Principal component analysis model of 16 q-FPs:</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>Images assessed with computerized image-analysis by two independent investigators to output the profile</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>of q-FPs data for each slice in operator-defined segmentation regions of liver tissue, including:</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>(1) General: the liver section in its entirety</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>(2) Perisinusoidal: hepatocyte-associated collagen in the perisinusoidal space</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>(3) Vessel: collagen fibrils directly connected to veins; and</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>(4) Vessel bridges: collagen fibrils extending from vein to vein or vein to portal tract.</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>70 q-FPs had interclass concordance ≥0.8 which were selected for further model development</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Wang et al. (2020)</td>
<td>q-FP</td>
<td>Compared against NASH CRN staging system (but with substages of stage 1 combined)</td>
<td>344 (428 biopsies)</td>
<td>25 q-FPs with AUC &gt;0.90 for different fibrosis stages; perimeter of collagen fibres and number of long collagen fibres had the best accuracy (88.3–96.2% sensitivity and 78.1–91.1% specificity for different fibrosis stages)</td>
</tr>
<tr>
<td>Chang et al. (2018)</td>
<td>SHG B-index</td>
<td>SHG/TPF to capture images (final sampling size of 10 mm² per biopsy)</td>
<td>83 adults</td>
<td>Prediction model based on 14 unique SHG-based collagen parameters</td>
</tr>
<tr>
<td></td>
<td></td>
<td>An image processing algorithm was used to quantify fibrosis features in three specific regions:</td>
<td></td>
<td>- Fibrosis vs. no fibrosis: AUC 0.853</td>
</tr>
<tr>
<td></td>
<td></td>
<td>1) central vein, 2) portal tract, and 3) perisinusoidal</td>
<td></td>
<td>- Cirrhosis vs. earlier stages: AUC 0.941</td>
</tr>
<tr>
<td></td>
<td></td>
<td>In total, 100 collagen features were extracted and quantified, of which 28 features including the percentages of different collagen patterns and collagen string features were extracted in each region</td>
<td></td>
<td>- Stage 0/1 vs. 2/3/4: AUC 0.967</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Compared against Brunt’s staging system</td>
<td></td>
<td>- Stage 0/1/2 vs. 3/4: AUC 0.985</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>- High correlation of 0.820 with fibrosis stage (P&lt;0.001)</td>
</tr>
<tr>
<td>Liu et al. (2017)</td>
<td>qFibrosis</td>
<td>SHG/TPF to capture images (final sampling size of 10 mm² per biopsy)</td>
<td>62 adults</td>
<td>Prediction model based on six shared parameters for string collagen</td>
</tr>
<tr>
<td></td>
<td></td>
<td>An image processing algorithm was used to quantify fibrosis features in three specific regions:</td>
<td>(30 training, 32 validation);</td>
<td>(Adult)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>1) central vein, 2) portal tract, and 3) perisinusoidal</td>
<td>36 children</td>
<td>- Fibrosis vs. no fibrosis: AUC 0.835</td>
</tr>
<tr>
<td></td>
<td></td>
<td>In total, 100 collagen features were extracted and quantified</td>
<td>(18 training, 18 validation)</td>
<td>- Cirrhosis vs. earlier stages: AUC 0.982</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>- Stage 0/1 vs. 2/3/4: AUC 0.892</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>- Stage 0/1/2 vs. 3/4: AUC 0.87</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>- Fibrosis vs. no fibrosis: AUC 0.981</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>- Stage 0/1 vs. 2/3: AUC 0.931</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>- Stage 0/1/2 vs. 3: AUC 0.885</td>
</tr>
<tr>
<td>Liu et al. (2020)</td>
<td>qFibrosis</td>
<td>Compared against NASH CRN staging system</td>
<td>219 adults</td>
<td>Prediction model based on 17 parameters, with output as a numerical index from 0 and 6.55</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>(146 training, 73 validation) (multicenter)</td>
<td>- Fibrosis vs. no fibrosis: AUC 0.87</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>- Cirrhosis vs. earlier stages: AUC 0.951</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>- Stage 0/1 vs. 2/3/4: AUC 0.881</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>- Stage 0/1/2 vs. 3/4: AUC 0.945</td>
</tr>
</tbody>
</table>

SHG, second harmonic generation; NAFLD, nonalcoholic fatty liver disease; q-FP, quantification of fibrosis-related parameter; TPEF, two-photon excitation fluorescence; AUC, area under curve; NASH CRN, Nonalcoholic Steatohepatitis Clinical Research Network.
(hepat)opathologists. The qFIBS (Fibrosis, Inflammation, Ballooning, and Steatosis) model, an SHG-based automated quantitative tool developed and validated by Liu et al.\(^7,8\) in a multicenter cohort, shows promise in mitigating these issues. This computational algorithm expands on the initial qFibrosis model developed in CHB patients; it quantifies the four key histopathological features of NASH, namely, fibrosis (qFibrosis), lobular inflammation (qInflammation), hepatocellular ballooning (qBallooning), and steatosis (qSteatosis). In their study, each qFIBS components were strongly correlated with the corresponding NASH CRN components (\(P<0.001\)). Validation of qFibrosis showed strong correlation with histological fibrosis stage (\(r=0.776\)) and accurate differentiation of fibrosis stages. Of note, however, is that similar to the NASH CRN scoring system, qFIBS is not suited for establishing a de novo diagnosis but is more of an adjunctive tool to quantify disease severity.

All these various models have affirmed SHG microscopy to be an invaluable new platform to study and quantify liver fibrosis, as well as the other key histological parameters of NASH. As current fibrosis staging systems tend to be disease-specific, the strong correlation between the models and traditional histological scoring systems used specifically in NAFLD promises great clinical utility, especially where demonstration of histological fibrosis improvement is an endpoint in drug trials for NAFLD. Similar to CPA measurements, SHG-based microscopy techniques allow for an objective quantitative assessment of fibrosis changes on a continuous scale, thereby providing greater reflection of subtle nuances compared to the stage migration of the traditional simpler semi-quantitative scores. However, in addition, combination with techniques such as TPEF or CARS allows for assessment of spatial information of collagen fibres, which are lacking in CPA measurements. In fact, the potential of qFibrosis could be further developed beyond the traditional confines of grouping or staging fibrosis as a single component, and instead be scored separately as the three subindices of perisinusoidal, portal, and septal fibrosis. This more refined fibrosis categorization would potentially be more sensitive and meaningful in monitoring patients’ disease progression. Regardless, it must be remembered that SHG microscopy still requires liver tissue samples to be performed on, and thus is subject to the sampling variances and the risks of an invasive biopsy procedure. The requirement for specialised equipment may also limit its utility in resource-poor areas or countries.

**ARTIFICIAL INTELLIGENCE (AI)-ASSISTED SYSTEMS**

Current computer-assisted SHG/TPEF image analytics for liver fibrosis scoring is not fully automated as it entails manual pre-processing (segmentation and feature extraction) based on domain knowledge in liver pathology, potentially introducing bias. For example, qFIBS uses an algorithm-derived platform rather than a machine-learning platform, resulting in correlations but not exact correspondence between the pertinent variables.\(^8\) Deep learning-based algorithms can potentially classify these images through learning from a large dataset of images. Yu et al.\(^8\) have shown that deep learning-based algorithms with transfer learning can automatically quantify liver fibrosis progression and score different stages of fibrosis (based on the METAVIR scoring system) with high sensitivity and specificity in a rat model. The level of accuracy by this fully automated algorithm using pre-trained AlexNet-Convolutional Neural Networks is similar to conventional non-deep learning-based algorithms in scoring liver fibrosis stages. However, this approach has yet to be validated in NAFLD patients.

A significant barrier to the widespread adoption of many of the newer technologies is the large computational effort and specialised equipment required. Recognising this difficulty, Forlano et al.\(^9\) developed a high-throughput, machine learning-based and fully automated method of quantification of steatosis, inflammation, ballooning, and collagen in liver biopsies from NAFLD patients that was user-friendly, fast-operating, and accurate. The devised software algorithm analysed biopsy images to compute percentages of each of the three key features of NASH, as well as calculated the CPA, with levels of inter- and intra-observer agreement ranging from 0.95 to 0.99, higher than that of semiquantitative scoring systems. It was also more sensitive in detecting differences in paired liver biopsies compared with the NASH CRN scoring system. The machine learning software is touted to be easily installable on any device and quantification performed within 2 minutes, features that would be attractive, especially in low resource and non-specialist centers.

Most recently, Gawrieh et al.\(^4\) also developed an integrated AI-based automated tool to detect and quantify liver fibrosis in NAFLD liver biopsies via CPA; however, in addition, their method also included assessment of six fibrosis architectural patterns. This added feature helps compensate for the absence of spatial information when using CPA alone for quantitative assessment of liver fibrosis, especially since the authors found considerable overlap in CPA across different stages.
It remains to be seen how well these AI-assisted tools perform alone in large scale validation studies. However, these exciting new developments also have the potential to be integrated with SHG-based microscopy to produce a model for assessing liver fibrosis in NAFLD which is sensitive and accurate, and yet also fully automated, widely accessible, and fast-operating.

**LIVER COLLAGEN FRACTIONAL SYNTHESIS RATE (FSR)**

Most of the above approaches are static measurements of fibrosis and do not give much information regarding the trajectory of disease progression. Decaris et al. recently proposed a novel method to quantify hepatic fibrogenesis flux rates both within liver tissue and noninvasively in blood, via administration of heavy water and tandem mass spectrometry to measure the liver collagen FSR and plasma lumican FSR. They found that the hepatic collagen FSR in NAFLD increased with advancing disease stage, suggesting that even at advanced stages, fibrosis may be reversible if ongoing collagen deposition rates can be reduced. Patients demonstrating a high FSR may thus benefit most from antifibrotic therapies.

**INVESTIGATING OTHER CONTRIBUTORS TO FIBROSIS**

**Elastin**

Fibrosis is a mixture of various ECM proteins and glycoproteins, of which collagen is among the most abundant but not the sole component. Elastin is actively synthesized by portal fibroblasts and possibly HSC in the diseased liver, and is biochemically stable compared with collagen. Nakayama et al. found that the presence of perivenular elastic fibers was associated with at least stage 3 fibrosis, and suggested that this may be a useful marker to determine advanced fibrosis in NAFLD; similarly, Masugi et al. detected higher elastin area ratios in advanced stages of NAFLD. Future studies may further clarify the clinical significance of elastin deposition with respect to disease reversibility as well as clinical outcomes.

**Activated hepatic stellate cells**

Activated HSC can be detected with antibodies against α-smooth muscle cells (ASMA). The degree of HSC activation correlates with the degree of fibrosis in NAFLD patients. Feldstein et al. devised an HSC activation score using a semiquantitative 12-point scoring system based on ASMA immunohistochemistry, and tested it in a cohort of 39 untreated NAFLD patients with paired liver biopsies. In their study, the HSC score was accurate in predicting fibrosis progression, suggesting that this score may be a useful adjunct to existing staging systems in predicting patients’ fibrosis progression rate, and in patient stratification in antifibrotic clinical trials.

**Liver sinusoidal endothelial cells (LSEC)**

LSEC are now known to be major effectors of inflammation in NASH and thereby contributors to fibrosis. Capillarization of LSECs and LSEC dysfunction appear to precede as well as promote liver fibrosis. Baiocchini et al. recently found that capillarization was only observed at initial stages of liver fibrosis in CHC patients. There are as yet no studies focused on correlating LSEC changes with different stages of fibrosis in NAFLD.

**Molecular pathways**

*In situ* mass spectrometry has been performed on liver biopsies, allowing direct characterization of the spatial distribution of various lipid species on tissue sections and providing insights into the pathogenesis of NASH and its progression. Urasaki et al. also used hyperspectral simulated Raman scattering microscopy and nanofluidic proteomics to quantitatively measure the liver composition of protein, DNA, and lipids and identify affected cellular signalling and metabolic pathways. However, these techniques have yet to be applied in investigating fibrosis stage in NAFLD.

**FUTURE DIRECTIONS**

There are many unmet needs in NAFLD. The pathogenesis of the disease and mechanisms responsible for liver fibrosis and remodeling in cirrhosis are still unchartered. The significance of the histological features required for the diagnosis of NASH, and the presently used efficacy endpoints in NASH clinical trials including
the absence of ballooning and at least 1-point decrease in fibrosis stage based on current histological scoring systems, may not be adequate and are being questioned.97

There have been many exciting recent developments and research pathways to address the issue of liver fibrosis. So far, despite its shortcomings, none of the noninvasive tools can currently replace liver biopsy as the reference standard in evaluating the various histological patterns of disease and their severity in NAFLD. The recent application of SHG microscopy on liver tissue samples promises to further refine the accuracy of this ‘gold’ standard, thereby changing the landscape of the role of the liver biopsy (Fig. 2).

SHG-based tools can provide a more standardized, accurate, and precise approach to staging NASH by incorporating a quantitative assessment of changes on a continuous scale across the stages of fibrosis, detecting subtle nuances and characteristics of the deposited collagen fibers, as well as characterizing spatial and architectural changes. SHG microscopy therefore outperforms current staging systems in capturing the full spectrum of fibrosis in NASH, and provides a more reproducible, tractable, and sensitive reference to analyse progression or regression of fibrosis in NASH. Current models can be further refined to assess the severity of perisinusoidal fibrosis at all stages of fibrosis. Integrating the use of AI-assisted systems to achieve a fully-automated and fast-operating system may also facilitate wider adoption of these models.

Much more work needs to be done before these proposed systems and models are used routinely in clinical practice and therapeutic trials. One thing, however, is certain: the liver tissue biopsy still has much to contribute to deepening our understanding and assessment of liver fibrosis in NAFLD.

Authors’ contributions
Gwyneth Soon and Aileen Wee contributed to the literature review and manuscript preparation.

Acknowledgements
The authors would like to thank Dr. Pang Yin Huei for critical review of the draft manuscript, and Dr. Dean Tai of HistolIndex for contributing the microscopy images.

Conflicts of Interest
The authors have no conflicts to disclose.

REFERENCES
8. Castera L. Non-invasive tests for liver fibrosis in NAFLD: creating
10. European Association for the Study of the Liver (EASL); European Association for the Study of Diabetes (EASD); European Association for the Study of Obesity (EASO). EASL-EASD-EASO clinical practice guidelines for the management of non-alcoholic fatty liver disease. J Hepatol 2016;64:1388-1402.


40. Pirola CJ, Sookoian S. Multiomics biomarkers for the prediction of nonalcoholic fatty liver disease severity. World J Gastroenterol 2018;24:1601-1615.


56. Bedossa P; FLIP Pathology Consortium. Utility and appropriateness of the fatty liver inhibition of progression (FLIP) algorithm and steatosis, activity, and fibrosis (SAF) score in the evaluation of biopsies of nonalcoholic fatty liver disease. Hepatology 2014;60:565-575.


